STRUCTURE OF STICHOPOGENIN A_L , THE GENUINE AGLYCONE OF HOLOTOXIN A ISOLATED FROM STICHOPUS JAPONICUS SELENKA

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Recently, we reported¹⁾ the elucidation of the carbohydrate portion of holotoxin A, a major antifungal glycoelde of Stichopua japonicus SBLRNKA, and also described the Identity of its triterpenoid aglycone with stichopogenin A_{Λ} , to which the structure $\frac{1}{\Lambda}$ was previously proposed by Elyakcv, et al.²⁾ The structure 1 was characteristic by the possession of a rare example of an unconjugated $\Delta^{5,8}$ -diene moiety and only limited numbers of sterols having the similar diene system have so far been isolated in the free states.³⁾ Since survival of the $\Delta^{5,8}$ -diene system during acid hydrolysis of the parent glycoside appeared significant and the diene system was expected to show a red shift In its CD spectrum due to the interaction of the **two** π **→** π **transitions⁴)** and in order to elucidate the genuine aglycone of holotoxin A, we have examined the CD spectra of holotoxin A and its aglycone (= stichopogenin A_{λ}). However, the spectra have revealed that both stichopogenin A_{λ} and holotoxin A possess unexpectedly the ketone chromophore along with the olefin and γ -lactone chromophores in their molecules (stichopogenin A_4 : [0]₂₀₀ +41000, [0]₂₃₃ -21000, [0]₃₀₅ -16800; holotoxin A: [0]₂₀₅ +22000, [0]₂₃₃ -13700, $[0]_{905}$ -9900). On the basis of the following evidence, we have reached the conclusion that stichopogenin A_A is expressed as 2 rather than 1 and consequently that the total structure of holotoxin A is formulated as 3 (the configuration at C-20 being undefined yet).

Stichopogenin A₄ (2), C₃₀H₄₆O₅'1/2H₂O, mp 240-243°, [a]_D²³-77° (CHCl₃), Mass (m/e): 486 (M^+) , 468 $(M^+ - H_2O)$, shows no UV absorption above 210 nm but a broad IR absorption band at 1750

 c_m^{-1} (KBr). On acetylation with Ac₂0/pyridine, 2 gave a monoacetate (2a), C₃₂H₄₈O₆, M⁺: m/e 528, mp 220-222° (250-252°⁵⁾), $[\alpha]_D^{23}$ -53° (CHCl₃), IR (CHCl₃): 1732 (br), 1716 (sh) cm⁻¹. These physical data are in accord with those reported for stichopogenin A_L and its monoacetate by Elyakov, et al. $^{2)}$

Since the CD spectrum of 2 shows the presence of the ketone, which does not locate in the 6-membered ring as revealed by the IR spectrum of 2 , 2 is considered to carry one double bond and one 5-membered ring ketone in place of two double bonds and one hydroxyl as seen in 1 . The ketone is assigned at C-16 in the lanostane skeleton as based on the negative CD curve ($\begin{bmatrix} \theta \end{bmatrix}_{305}$ -16800: cf. $6 - 10^{6c}$). The assignment is further supported by a fact that the addition of alkali alters the CD curve of 2 to a curve ($\left[\theta\right]_{270}$ -22400, $\left[\theta\right]_{330}$ -5300 (sh)) ascribable to an enone chromophore (4) .⁷⁾ Therefore, the IR absorption band due to the C-16 ketone in 2 seems to be overlapped by the band due to the Y-lactone (vide infra).

In the PMR spectrum of stichopogenin A_4 (2), a one-proton signal (br, $W_{h/2}$ = 6 Hz) is observed at 65.27 and hence the double bond could be assigned as Δ^5 , Δ^7 , or $\Delta^{9(11)}$. Among them, $\triangle^{9(11)}$ is preferred, since the observed strong positive CD maximum ($\left[\theta\right]_{200}$ +41000) is in good accord with the reported values for the $\Delta^{9(11)}$ -lanostene triterpenoids (cf. μ - μ^{8}). addition, the olefinic proton chemical shift is quite alike to those reported for the $\Delta^{9(11)}$ lanostene derivatives.⁹⁾

As for the γ -lactone moiety of stichopogenin A_4 (2), the n+ π transition is observed with the strong negative CD maximum ($\left[\theta\right]_{233}$ -21000). The sign is well explained by the Beecham's rule,¹⁰⁾ and the red shift and the enhanced magnitude could be attributable to the spacial interaction of the $\Delta^{9(11)}$ double bond located near the γ -lactone moiety.¹¹⁾ The explanation is also supported by comparing the CD data of 2 with those of the $9a,11a$ -epoxy derivative (5) $(\left[0\right]_{228}$ -5400, $\left[0\right]_{305}$ -12000), which was prepared from 2 by m-chloroperbenzoic acid oxidation.

In the structural study by Elyakov, et al.²⁾ who proposed the structure 1 for stichopogenin A_{μ} , 17a-OH in 1 was based on the angular CH₃ chemical shifts which were assigned in comparison with those of 22,25-oxido-holothurinogenin (16). However, since 10-CH₃'s in the $\Delta^{9(11)}$ -lanostene triterpenoids are also observed at the similar positions $^{7a,12)}$, the above assignment could not be the definite criterion. Furthermore, the following evidence excludes the presence of 17 α -OH in stichopogenin A₄ (2).

As described previously, ¹⁾ the mild acid hydrolysis (aq. 2% $H_2SO_4/MeOH/b$ enzene) of holotoxin A (2) furnished 2 and another aglycone (2b), $C_{31}H_{48}O_5$, mp 240-243°, [a] $^{24}_{D}$ -96° (MeOH); Mass(m/e)

: 500 (M^+) , 468 $(M^+$ -CH₃OH), 73 ($(CH_3)^2$ C $=$ OCH₃)¹³; PMR (CDC1₃, 6): 0.84, 0.89, 0.99, 1.19 (3H each, all s), 1.12 (6H, s), 1.41 (3H, s)(CH₃x7), 3.17 (3H, s, CH₃0), 3.00-3.33 (2H, m. 3a-H, 86-H), 5.29 (1H, m, 11-H). The CD spectrum of 2b shows the presence of the same chromophore as in 2 (${[6]}_{200}$ +33000, ${[6]}_{233}$ -16400, ${[6]}_{303.5}$ -15500). Acetylation of 2b with Ac₂0/pyridine gave a monoacetate (2c), mp 238-241°, ${a \choose b}$ -11° (CHC1₃), the IR spectrum (CC1₄) of which shows no OR absorption band, but the absorption bands due to y-lactone, 5-membered ring ketone, and acetoxyl (1768 (sh), 1751, 1742 (sh) cm^{-1}).

Stichopogenin A₄ (2) is considered to be a genuine aglycone of holotoxin A (3), since i) 2 is obtained by the mild acid hydrolysis, and ii) the CD spectra of both 2 and 3 show the presence of the same chromophores in both compounds, although the maxima intensities in the CD spectrum of $\frac{3}{2}$ are slightly lower than in $\frac{2}{2}$, the reason of which will be a subject of the further investigation.

References and Footnotes

- 1) I. Kitagawa, T. Sugawara, and I. Yosioka, <u>Tetrahedron Letters, 1974</u>, 4111.
- 2) G. B. Elyakov, T. A. Kuzunetsova, A. K. Dzizenko, and Y. K. Elkin, <u>Tetrahedron Letters,</u> 1969, 1151.
- 3) a) N. Sugiyama, M. Yamamoto, and K. Yamada, <u>Mippon Kagaku Zaeshi, 89</u>, 710 (1968); b) T. 1 Lenton, L. J. Goad, and T. W. Goodwin, Phytochemistry, 12, 1135 (1973).
- 4) G. Snatzke ed., "Optical Rotatory Dispersion and Circular Dichroism in Organic Chemistry", Heyden 6 Son Ltd., London, 1967, p. 153.
- 5) Double malting-point.
- 6) a) P. Crabbé, "ORD and CD in Chemistry and Biochemistry", Academic Press, New York, 1972, p. 35; b) L. Velluz and M. Legrand, Angew. Chem., 73, 603 (1961); c) K. Kuriyama, unpublished data.
- 7) a) I. Rothberg, B. Tursch, and C. Djerassi, <u>J. Org. Chem</u>., 38, 209 (1973); b) C. Djerassi, R. Records, E. Bunnenberg, K. Mislow, and A. Moscowitz, J. Am. Chem. Soc., 84, 870 (1962).
- 8) a) H. Irie, S. Uyeo, and K. Kuriyama, <u>Tetrahedron Letters, 1971</u>, 3467; b) A. I. Scott and A. D. Wrixon, Tetrahedron, 26, 3695 (1970).
- 9) a) J. P. Kutney, D. S. Grierson, G. D. Knowlis, and N. D. Westcott, <u>Tetrahedron, 29</u>, 13 (1973); b) S. Uyeo, J. Okada, S. Matsunaga, and J. W. Rowe, Tetrahedron, 24, 2859 (1968).
- 10) A. F. Beecham, Tetrahedron Letters, 1968, 2355, 3591.
- 11) a) H. Labhart and G. Wagniere, $Relv. Chim. Acta, $\frac{42}{2}$, 2219 (1959); b) R. C. Cookson and S.$ </u> Mackengie, Proc. Chem. Soc., 1961, 423; c) A. Moscowitz, K. Mislow, M. A. W. Glass, and C. Djerassi, J. Am. Chem. Soc., 84, 1945 (1962).
- 12) A. I. Cohen, D. Rosenthal, G. W. Krakower, and J. Fried, Tetrahedron, 21, 3171 (1965).
- 13) a) R. Roller, C. Djerassi, R. Cloetens, and B. Tursch, J. Am. Chem. Soc., 91, 4918 (1969);b) G. Habermehl and G. Volkwein, Ann., 731, 53 (1970).